

Mangiferin Trimethyl Ether—A suspension of mangiferin (30 mg.) in 2 ml. methanol and 8 ml. ether was treated with an excess of ethereal diazomethane following the procedure of Hawthorne and co-workers (7). After twice being precipitated from aqueous acetone, the product had m.p. 298–299° and provided infrared and ultraviolet spectra identical with those of an authentic sample. Literature m.p. 298–299° (7).

REFERENCES

- (1) Finnegan, R. A., and Mueller, W. H., *J. Pharm. Sci.*, **54**, 1136(1965).
- (2) Gorter, K., *Bull. Jard. Bot. Buitenzorg (III)*, **2**, 187(1920).
- (3) Finnegan, R. A., and Stephani, R. A., to be published.
- (4) Roberts, J. C., *Chem. Rev.*, **61**, 591(1961).
- (5) Finnegan, R. A., Patel, J. K., and Bachman, P. L., *Tetrahedron Letters*, (1966)6087.
- (6) Iseda, S., *J. Chem. Soc. Japan*, **30**, 625, 629(1957).
- (7) Hawthorne, B. J., Janes, N. F., King, F. E., and Morgan, J. W. W., "Recent Progress in the Chemistry of Natural and Synthetic Colouring Matters and Related Fields," Gore, T. S., ed., Academic Press Inc., New York, N. Y., 1962, p. 331.
- (8) Haynes, L. J., and Taylor, D. R., *J. Chem. Soc.(C)*, (1966)1685.
- (9) Gorter, K., *Bull. Jard. Bot. Buitenzorg (III)*, **4**, 260(1922); through *Chem. Abstr.*, **17**, 1472(1923).
- (10) Paris, R., and Etchepare, S., *Compt. Rend.*, **258**, 5277(1964).
- (11) Haynes, L. J., *Adv. Carbohydrate Chem.*, **18**, 227(1963); **20**, 357(1965).
- (12) Ramanathan, J. D., and Seshadri, T. R., *Current Sci.*, **29**, 131(1960).
- (13) Pillay, P. P., and Lekshmi, A., *Bull. Res. Instit. Univ. Kerala Trivandrum*, **5**, 47(1957); through *Chem. Abstr.*, **52**, 20423h(1958).
- (14) Horhammer, L., and Wagner, H., "Recent Developments in the Chemistry of Natural Phenolic Compounds," Ollis, W. D., ed., Pergamon Press, New York, N. Y., 1961, p. 185.
- (15) Bate-Smith, E. C., and Harborne, J. B., *Nature*, **198**, 1307(1963).
- (16) Ueno, A., *Yakugaku Zasshi*, **82**, 1482(1962).
- (17) Subramanian, S. S., and Nair, A. G. R., *Ind. J. Chem.*, **4**, 335(1966).
- (18) Morita, N., Shimizu, M., and Fukuta, M., *Yakugaku Zasshi*, **85**, 374(1965).
- (19) Andriantsiferana, R., *Compt. Rend. Ser. D*, **264**, 1215(1967).
- (20) Andriantsiferana, B., *Compt. Rend. Soc. Biol.*, **159**, 1899(1965).
- (21) Finnegan, R. A., Merkel, K. E., Patel, J. K., and Back, N., presented to the Medicinal Chemistry Section, APHA Academy of Pharmaceutical Sciences, 114th Annual Meeting, Las Vegas, Nevada, April 1967, Abstracts of Papers, p. 77.
- (22) Nirenberg, M., Matthaei, J., Jones, O., *Proc. Natl. Acad. Sci.*, **48**, 104(1962).
- (23) Coutsoygeorgopoulos, C., *Biochim. Biophys. Res. Commun.*, **27**, 46(1967).



Keyphrases

Hiptage madablota
Mangiferin— isolation, identification
Biological activity— mangiferin
IR spectrophotometry— structure
UV spectrophotometry— structure

Deficiency of Vitamin B₁₂ in *Chlorella*

By ROBERTSON PRATT and EVELYN JOHNSON*

Repeated systematic attempts to detect vitamin B₁₂ in two species of *Chlorella* at different times during the culture growth period failed to reveal consistent or significant evidence of B₁₂ or B₁₂-like activity. The results are discussed in terms of other reports in the literature.

AS PART of a continuing investigation of the vitamin content of two *Chlorella* species (1–4), extracts of *C. vulgaris* and of *C. pyrenoidosa* have been assayed for vitamin B₁₂ (cyanocobalamin and related factors). This seemed of particular interest in view of the conflicting reports in the literature concerning occurrence of the vitamin in chlorophyllous plants—especially the algae and particularly the Chlorococcales.

In an important review of vitamin B₁₂ in the metabolism of microorganisms, Ford and Hutner (5) state "B₁₂ . . . is not present in green plants and yeast; it is synthesized by bacteria and probably by blue-green, brown, and red algae, but not by green algae." Others have suggested that B₁₂ found associated with seaweeds (whether red, brown, or green) can be attributed to bacteria living epiphytically on the algae (6, 7) or to bacterial contami-

nants in the distilled water used in preparing specimens for assay (8).

Referring specifically to *Chlorella* (species unspecified), Hutner (9) commented on the lack of B₁₂ in the organism while, in contrast, Hashimoto reported significant levels of the vitamin (av. 61 μ mcg./mg. dry wt.) in "pure cultured" *Chlorella ellipsoidea* and he considered this a sufficiently high level to warrant further study (10).

METHODS

Chlorella vulgaris and *C. pyrenoidosa* were cultured aseptically by both the column and the round-bottom flask techniques described previously (11): light intensity in the former was 600 ftc. and in the latter 1250 ftc. Column cultures were harvested at different times from Day 5 to Day 21; flask cultures were harvested at intervals from 3 weeks to 10 weeks postinoculation.

Assays were performed using *Euglena gracilis* (ATCC 12716) and *Ochromonas malhamensis* (ATCC 11532) in the appropriate Difco media. Acid-cleaned glassware and fresh glass-redistilled

Received November 13, 1967, from the School of Pharmacy, University of California, San Francisco Medical Center, San Francisco, CA 94122

Accepted for publication January 18, 1968.

* Present address: Department of Biology, University of California at San Diego, La Jolla, California.

water were used for all assays. Standard curves were determined in each assay using USP cyanocobalamin reference standard.

Three basic extraction procedures were employed: (a) water extraction with heat, (b) refluxing with 70% isopropanol, and (c) extraction with perchloric acid. A number of variations of these were tried including (a) addition of sodium cyanide (10 mg./ml.), (b) buffering with 0.01 N sodium acetate or adjusting pH with HCl, (c) heating at 90° or autoclaving, (d) subsequent digestion with papain, and (e) preliminary mincing in a mortar with crystalon abrasive.

No single procedure seemed consistently or distinctly superior for releasing B₁₂ (if present) from the algae. In some instances extracts prepared with sodium acetate seemed to be slightly inhibitory for the *Euglena* test organism; use of isopropanol resulted in an extract containing lipids and other cell material that interfered with turbidity readings during actual assay; the perchloric acid technique resulted in extracts significantly inhibitory to growth of *Euglena*, due to inability to completely remove the perchloric acid, and the papain technique gave a high blank reading that overshadowed any possible B₁₂ in *Chlorella*.

Therefore, the method recommended by the Analytical Methods Committee (12) was used routinely. In this procedure a suspension of cells with 0.01% NaCN (pH 4.5 with HCl) was heated in a water bath at 90° for 30 min. Recovery of added B₁₂ was approximately 110%.

RESULTS

In seven different experiments between October 1966 and August 1967, each entailing several assays of extracts prepared from one to five harvests made at different times during the culture period, no significant or consistent levels of B₁₂ or B₁₂-like activity were detected. Most frequently no B₁₂ was found. Low levels (0.1 to 5 μmcg./mg. dry wt.) were found sporadically, generally in cultures 14 days or more postinoculation.

Although aseptic techniques are employed in setting up the algal cultures, the danger of bacterial contamination having occurred during subsequent manipulation cannot be totally eliminated, especially for older cultures. Therefore, the possibility of low-

level bacterial contamination as the cultures age must be considered as a possible source of B₁₂.

Moreover, the B₁₂ levels found were generally so low that often they were within the range of error inherent in the extraction and assay procedures.

The possibility exists that extraction procedures were not adequate for releasing B₁₂ from the *Chlorella* cells but, in view of the variety of procedures and modifications employed, it seems unlikely that all of the methods would have been totally inadequate. Nonetheless, the possibility that B₁₂ is protein bound and not readily released or that the tough *Chlorella* cell wall prevents release should not be overlooked.

Other possibilities are (a) the synthetic medium employed for culturing the algae precluded or interfered with biosynthesis of the vitamin, and/or (b) B₁₂ is synthesized but so quickly transferred in one or more various synthetic pathways that it never accumulates in assayable levels.

However, under the conditions employed for culture and for the variety of techniques for extracting the cells and assaying the extracts, the authors have been unable to demonstrate the presence of B₁₂ or B₁₂-like activity in *Chlorella vulgaris* and *C. pyrenoidosa*.

REFERENCES

- (1) Pratt, R., and Johnson, E., *J. Pharm. Sci.*, **53**, 151 (1964).
- (2) *Ibid.*, **54**, 871 (1965).
- (3) *Ibid.*, **55**, 799 (1966).
- (4) *Ibid.*, **56**, 536 (1967).
- (5) Ford, J. E., and Hutner, S. H., in "Vitamins and Hormones," Harris, R. E. ed., Academic Press, Inc., New York, N. Y., 13, 101, 1955.
- (6) Ericson, L. E., and Lewis, L., *Arkiv. Kemi.*, **6**, 427 (1953).
- (7) Robins, W. J., Hervey, A., and Stebbins, M. E., *Bull. Torrey Bot. Club*, **78**, 363 (1951).
- (8) Robins, W. J., Hervey, A., and Stebbins, M. E., *Ann. N. Y. Acad. Sci.*, **56**, 818 (1953).
- (9) Hutner, S. H., *Protozoan*, **2**, 13 (1955).
- (10) Hashimoto, Y., *J. Vitaminol.*, **1**, 49 (1954).
- (11) Pratt, R., and Johnson, E., *J. Pharm. Sci.*, **52**, 975 (1963).
- (12) Analytical Methods Committee, *Analyst*, **81**, 132 (1956).



Keyphrases

Vitamin B₁₂

Chlorella species—vitamin B₁₂ content

Cultures—column, round-bottom flask techniques